

Incorporation of C¹⁴-labeled AMP into ATP is the most direct evidence for reversibility of amino acid incorporation into RNA. No incorporation of AMP into ATP occurred when RNA plus free amino acid was used in place of amino acid-RNA, or when P-P was omitted. Calculation of the data shows that the molar rate of AMP incorporation into ATP greatly exceeds the rate of amino acid-RNA breakdown, suggesting that reactions which form unlabeled ATP occur during the incubation.

The approximate equilibrium position of these reactions was near unity, indicating the high energy nature of the amino acid-RNA linkage.

(8) Work done during the tenure of an Established Investigatorship of the American Heart Association. These studies were supported by a grant from the National Science Foundation.

BIOCHEMISTRY DEPARTMENT
MEDICAL RESEARCH INSTITUTE
CITY OF HOPE MEDICAL CENTER
DUARTE, CALIFORNIA

EDWARD GLASSMAN
ESTHER H. ALLEN
RICHARD S. SCHWEET⁵

RECEIVED JUNE 2, 1958

16-ALKYLATED CORTICOID. I. 16 α -METHYL-PREDNISONE AND 16 β -METHYLPREDNISONE¹

Sir:

The recent publication by Boland² on 16 α -methyl corticosteroids prompts us to report our studies of both 16 α - and 16 β -methyl steroids which possess a high order of anti-inflammatory activity without salt retention in animal and clinical trials.

Reaction of 3 α -acetoxy-16-pregnene-11,20-dione³ (I) with diazomethane gave an intermediate pyrazoline,⁴ m.p. 199–200° dec., [α]_D +149.6° (all rotations 1% in diox.). *Anal.* Found: C, 69.51; H, 7.98; N, 6.69. Pyrolysis of this product at its melting point gave 3 α -acetoxy-16-methyl-16-pregnene-11,20-dione, m.p. 163–166°, [α]_D +69.9°, $\lambda_{\text{max}}^{\text{MeOH}}$ 248 m μ (ϵ 10,800). *Anal.* Found: C, 74.58; H, 8.55. Reduction with palladium yielded 3 α -acetoxy-16 β -methyl pregnane-11,20-dione (II) m.p. 160–163°, [α]_D +93.6°, no ultraviolet absorption between 220–300 m μ . *Anal.* Found: C, 74.37; H, 9.06. Enol acetylation with *p*-toluenesulfonic acid and acetic anhydride, then treatment with peracetic acid and finally alkaline hydrolysis, gave 3 α ,17 α -dihydroxy-16 β -methylpregnane-11,20-dione, m.p. 182–185°, [α]_D +83.6°. *Anal.* Found: C, 72.82; H, 8.92. Bromination at C-21 and then treatment with potassium acetate gave 3 α ,17 α -21-trihydroxy-16 β -methylpregnane-11,20-dione 21-acetate, m.p. 200–206°, [α]_D +119.6°. *Anal.* Found: C, 68.79; H, 8.39. Oxidation with N-bromosuccinimide produced the 3-ketone, m.p. 198–202°, [α]_D +128.0°. *Anal.* Found: C, 69.04; H, 8.10. Dibromination at positions 2 and 4, followed by dehydrobromination with dimethylformamide, produced 16 β -methylprednisone 21-acetate (III) m.p. 232–235°, [α]_D +213.6°.

(1) After submission of this manuscript, a Communication appeared [G. Arth, D. Johnston, J. Fried, W. Spooner, D. Hoff and L. Saret, *THIS JOURNAL*, **80**, 3180 (1958)] describing the preparation of 16 α -methylprednisone by essentially the same route. We have tried to eliminate as much of the common material as possible.

(2) E. W. Boland, *Cal. Med.*, **88**, 417 (1958).

(3) H. Slaters and N. Wendler, *J. Org. Chem.*, **22**, 498 (1957).

(4) Cf. A. Wettstein, *Helv. Chim. Acta*, **27**, 1803 (1944).

$\lambda_{\text{max}}^{\text{MeOH}}$ 238 m μ (ϵ 14,200). *Anal.* Found: C, 69.24, H, 7.21. Hydrolysis with potassium bicarbonate gave 16 β -methylprednisone, m.p. 210–204°, [α]_D +190.2°, $\lambda_{\text{max}}^{\text{MeOH}}$ 238 m μ (ϵ 14,700). *Anal.* Found: C, 71.19; H, 7.37

Reaction of I with methylmagnesium iodide produced 3 α -hydroxy-16 α -methylpregnane-11,20-dione,⁵ m.p. 149–154°, [α]_D +100.5°, no selective absorption between 200 and 340 m μ . *Anal.* Found: C, 74.16; H, 9.41. This was converted into 16 α -methylprednisone 21-acetate (m.p. 212–214° [α]_D +157.8°, $\lambda_{\text{max}}^{\text{MeOH}}$ 238 m μ (ϵ 15,500). *Anal.* Found: C, 69.84; H, 7.22) by the same procedure used for the conversion of II to III (*i.e.*, enol acetylation and peroxidation, 21-bromination and acetoxylation, oxidation at C-3 and 2,4-dibromination and dehydrobromination).

A direct comparison of prednisone and its 16-methyl derivatives in human subjects utilizing (1) metabolic balance studies⁶ consisting of the analysis of diet, urine and feces for calcium, phosphorus, nitrogen, sodium and potassium, and (2) the clinical response of patients indicate that 16-methylation (α or β) of the parent steroid, prednisone, is associated with an enhancement of anti-anabolic properties and an increase of 30–50% in both anti-inflammatory and sodium excreting properties. Unlike 16 α -hydroxylation, 16 α - or 16 β -methylation contributes to anti-inflammatory potentiation. The *in vivo* conversion of the 16-methyl corticoids into urinary 17-keto steroids is limited to less than 5%: a conversion slightly less than that obtained with the parent steroids unsubstituted at position 16, and much less than that obtained with cortisone or hydrocortisone.

(5) Cf. R. Marker and H. Crooks, *THIS JOURNAL*, **64**, 1280 (1942).

(6) E. C. Reifstein, P. Albright and S. Wells, *J. Clin. Endocrinol.*, **5**, 367 (1945).

EUGENE P. OLIVETO
RICHARD RAUSSER
A. L. NUSSBAUM
WILLIAM GEBERT
E. B. HERSHBERG
S. TOLKSDORF
MILTON EISLER
P. L. PERLMAN

RESEARCH LABORATORIES
SCHERING CORP.
BLOOMFIELD, N. J.

MASSACHUSETTS GENERAL HOSPITAL
BOSTON, MASS.

M. M. PECHET

RECEIVED JUNE 30, 1958

THE INTERMEDIATE COBALT HYDROCARBONYL-OLEFIN COMPLEX IN THE OXO REACTION¹

Sir:

The several mechanisms^{2,3,4} which have been suggested for the oxo synthesis all involve a rate-determining displacement of a mole of carbon monoxide from a carbonyl of cobalt by the attacking olefin. The present study shows that not only does complexing occur between olefin and hydrocarbonyl⁵ under room conditions without the libera-

(1) We wish to thank the Houdry Process Corp. for a generous fellowship which made this work possible.

(2) H. W. Sternberg, R. Markby and I. Wender, *THIS JOURNAL*, **79**, 6116 (1957); I. Wender and M. Orchin, in "Catalysis," Vol. V, Reinhold Publishing Corp., New York, N. Y. 1957, p. 124.

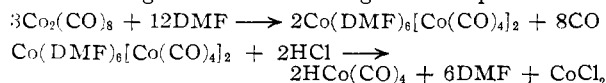
(3) A. R. Martin, *Chem. and Ind.*, 1536 (1954).

(4) G. Natta, R. Ercoli, S. Castellano and P. H. Barbieri, *THIS JOURNAL*, **76**, 4094 (1954).

(5) M. Orchin, L. Kirch and I. Goldfarb, *ibid.*, **78**, 5450 (1956).

tion of carbon monoxide but that the mixture instead absorbs carbon monoxide and complex decomposition leads to oxo aldehydes.

Dilute solutions of cobalt hydrocarbonyl were prepared from a toluene solution of dicobalt octacarbonyl at 25° under one atmosphere of carbon monoxide by disproportionation⁶ with dimethylformamide (DMF) and subsequent acidification of the resulting mixture according to the equations



Addition of olefin to the hydrocarbonyl solution caused carbon monoxide absorption and resulted in the gradual disappearance of the hydrocarbonyl as measured by *o*-phenanthroline.⁷ The relative rates of complex formation with 1-hexene, 2-hexene, and cyclohexene were 13:6:1. The relative rates of hydroformylation of these olefins at 110° are,⁸ respectively, 11.4:3.1:1.

At 0° excess 1-hexene reacts with hydrocarbonyl and carbon monoxide to give a stable complex. At 25°, the complex begins to decompose with evolution of carbon monoxide and formation of aldehyde. Because the complex with 2-hexene decomposes at a faster rate, it is unlikely that the two isomers form the same complex.⁹

In other experiments, the acid-dimethylformamide phase was removed prior to olefin addition. Addition of excess olefin resulted in the rapid absorption of one mole of carbon monoxide per two moles of hydrocarbonyl. The composition of the complex is unquestionably 2HCo(CO)₄·olefin·CO.

Preparation of Hydrocarbonyl and Reaction with 1-Hexene.—To a flask thermostated at 25° and connected to a gas buret,¹⁰ there was added 25 ml. of toluene and 1 ml. of dimethylformamide. After flushing with carbon monoxide, 5 ml. of toluene containing 0.51 g. (15 mmoles) of dicobalt octacarbonyl was added with stirring. Carbon monoxide liberation commenced immediately and, after one hour, 4 mmoles had been collected. Then 4 ml. (30 mmoles) of 1-hexene was injected through a serum-stoppered side-arm and the flask cooled to 0°. Two ml. of concentrated hydrochloric acid was injected and the mixture kept at 0° for 40 hours. Analysis (*o*-phenanthroline) showed that all the hydrocarbonyl had disappeared. In a blank experiment in the absence of olefin, 1.8 mmoles of hydrocarbonyl was present.

Carbon Monoxide Absorption.—Disproportionation of 3 mmoles of octacarbonyl was carried out at 25° as described above. The acid dimethylformamide phase was removed and 10 ml. (80 mmoles) of 1-hexene injected. Carbon monoxide absorption commenced immediately and was complete when two mmoles of gas was absorbed.

Aldehyde Formation.—In a disproportionation experiment using 10 ml. of dimethylformamide

and 90 ml. of toluene containing 27 mmoles of octacarbonyl, 72 mmoles of gas was liberated. Then 12 ml. of 1-pentene and 20 ml. of hydrochloric acid were injected. Gas evolution began at once. After standing overnight, the reaction mixture was poured into an alcoholic solution of 2,4-dinitrophenylhydrazine. The toluene was removed by distillation and the precipitated hydrazone recrystallized from ethanol, 2 g., m.p. 103–104°; literature¹¹ for hexaldehyde, 103°. *Anal.* Calcd. for C₁₂H₁₆N₄O₄: C, 51.45; H, 5.76; N, 20.00. Found:¹² C, 51.59; H, 5.67; N, 19.99.

(11) R. L. Shriner, R. C. Fuson and D. Y. Curtin, "Systematic Identification of Organic Compounds," 4th Ed., John Wiley and Sons, New York, N. Y., p. 283, 1956.

(12) Geller Laboratories, Hackensack, N. J.

DEPARTMENT OF CHEMISTRY
UNIVERSITY OF CINCINNATI
CINCINNATI 21, OHIO

LAWRENCE KIRCH
MILTON ORCHIN

RECEIVED JUNE 2, 1958

CRYSTALLINE HUMAN GROWTH HORMONE

Sir:

We wish to report methods for the isolation in high yields of crystalline human pituitary growth hormone. Li, *et al.*,¹ Ehrenberg and Heijenskjöld² and Raben³ also have recorded the preparation of human growth hormone, but not in crystalline form.

An acetone powder prepared from whole human pituitary glands was extracted with 0.3 *M* KCl at pH 5.5. The extract was brought to pH 8.5 and fractionated with ethanol in the cold,⁴ material precipitated by ethanol in the concentration range of 10–30% being collected. This was dissolved in 0.1 *M* KCl at pH 7.5 and ethanol added to 25% concentration. During addition of ethanol, a crop of impure crystals of growth hormone formed and was retained for subsequent purification. The supernatant liquor when refractionated yielded essentially homogeneous crystals. Amounts of hormone equal to or exceeding those obtained during the acid extraction were isolated by re-extracting the acetone powder residue with 0.3 *M* KCl at pH 11, and then ethanol fractionation.

Column chromatography with DEAE (diethylaminoethyl)-cellulose was used for final purification. Ellis and Simpson⁵ have used this exchanger with the beef hormone, but under different conditions. In our experiments the protein sample was applied to the column in pH 10, 0.02 *M* carbonate buffer, and the components were resolved by elution with buffer containing increasing amounts of sodium chloride. The human hormone was eluted at a salt concentration of 0.1–0.2 *M*. The columns were useful in recovering growth hormone from side fractions of the ethanol procedure. The hormone also has been obtained directly by

(6) I. Wender, H. Sternberg and M. Orchin, *THIS JOURNAL*, **74**, 1216 (1952).

(7) H. W. Sternberg, I. Wender and M. Orchin, *Anal. Chem.*, **24**, 174 (1952).

(8) I. Wender, S. Metlin, S. Ergun, H. W. Sternberg and H. Greenfeld, *THIS JOURNAL*, **78**, 5401 (1956).

(9) I. Goldfarb and M. Orchin, in "Advances in Catalysis," Vol. IX, Academic Press, Inc., New York, N. Y., 1957, p. 616.

(10) M. Orchin and I. Wender, *Anal. Chem.*, **21**, 875 (1949).

(1) C. H. Li and H. Papkoff, *Science*, **124**, 1293 (1956); Li, *Fed. Proc.*, **16**, 775 (1957).

(2) A. Ehrenberg and F. Heijenskjöld, *Acta. Chem. Scand.*, **10**, 1675 (1956).

(3) M. S. Raben, *Science*, **125**, 883 (1957).

(4) A. E. Wilhelmi, J. B. Fishman and J. A. Russell, *J. Biol. Chem.*, **176**, 735 (1948); E. Reid and A. E. Wilhelmi, *Proc. Soc. Expt. Biol. Med.*, **91**, 267 (1956).

(5) S. Ellis and M. E. Simpson, *J. Biol. Chem.*, **220**, 939 (1956).